BROMELIAD SOCIETY OF QUEENSLAND INC

P.O. Box 565,
FORTITUDE VALLEY,
QLD. 4006
AUSTRALIA

General meetings are held on the third Thursday of each month except December at the Uniting Church Hall, 52 Merthyr Road, New Farm, Queensland, commencing at 7.30 pm.

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Mr. Peter Paroz, Mr. Bert Wilson,

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'Bromeliaceae' is the Journal of the Bromeliad Society of Queensland Inc, and is published bimonthly.

PRINT POST NUMBER: P.P.434327/0002

VOL XXVIII Number 5 Price: $3.50

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Application should be in writing, should include all required copy for the advertisement, and should include payment by cash or cheque.

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Copy may be updated at any time if supplied in writing, otherwise last available copy will be used.

It is regretted that no photographs can be included in advertisements at this time.

The Society reserves the right to refuse publication of any advertisement.

Publication Deadlines for 'Bromeliaceae'

All articles, competition results, programme details and other items of interest for publication must be in the hands of the Editorial Committee by the following dates -

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EDITORIAL
John Higgins

It is a good thing, from time to time, to reflect upon the reasons that caused us to join the Society. While these reasons may be very diverse for many of us they all revolve around our interest in Bromeliads. The founding members of the Society no doubt shared very similar reasons for starting the Society. They also shared a love of these plants with one another and this serves as a common bond between the founders and our members of today. Similarly we will share this bond with the members of the future.

In their wisdom the founders produced a constitution to serve as a common guideline for the future of the Society. This constitution is an important link between the past, the present, and the future and provides us with a plan for growth and development. Minor amendments have been necessary to maintain the relevance of the constitution as changes have occurred in the state of the Society. The last amendments were made to enable the constitution to meet the requirements of Incorporation.

The Objects of the Society are at the very heart of our constitution and they are, in effect, a compact blueprint for the development and continued existence of the Society. One of these Objects is concerned with the development and promotion of bromeliads at large. This can be seen in action when we conduct public displays which provide a showcase of our plants to the public. Similarly, when we sell or give plants to others, this provides, through the enjoyment of ownership, an encouragement to expand involvement in bromeliads. All of this is based on the simple principle that the more people there are growing bromeliads the more likely it is that the plants will survive the depletion of habitats and the threat of extinction.

This is a heavy responsibility but is one that we all accept when we take on Society membership. One simple way we can all contribute to this important Object is to ensure that the labels we place on our plants are as accurate as possible. Future generations of growers will rely on what we bequeath them. This legacy should be positive. Simply expanding the written name on a plant label from ‘N. correia-araufi’ to ‘Neoregelia (or Nidularium!) correia-araufi’ may eliminate some confusion and preserve an important genetic identity for the future.
Cover Photograph - Front

_Aechmea fasciata_
First classified in 1828 as _Billbergia fasciata_, and reclassified as _Aechmea fasciata_ by T.G. Baker in 1879. A beautifully proportioned vase-like plant, usually with grey-green leaves barred with silver cross-bands. A striking pink pyramidal inflorescence with lavender-blue flowers ensure this distinctive and colourful plant remains one of the most popular of all bromeliads. This species originates from the Atlantic forests of Brazil where it grows epiphytically at elevations ranging from sea level to 1200m or more. It varies considerably in appearance according to the particular region in which it is found. Some forms have almost totally green leaves, others slender, narrow leaves with dense white bands and near red inflorescence. Still others are found with wide, white scaled leaves and pink inflorescence.

The photographed plant is one of the many cultivars of the species which have been developed over decades of cultivation. Its grey-green leaves with distinctive barring and bright pink inflorescence are a very attractive combination.

GROWN BY: Don & Phyl Hobbs  
PHOTOGRAPH BY: Don Hobbs

Cover Photograph - Back
_Vriesea lutheriana (J.R. Grant)_

A large, stemless, recently discovered species with ornamental appeal, _V. lutheriana_ has only just entered cultivation. Occurring locally, but sparsely, in the middle elevation rainforests of southeastern Costa Rica, it has probably only been collected once by Chester Skotak, those few plants being the source of the little cultivated material. It is a handsome and hardy plant, growing into a dense rosette of thick, arching, deep green, densely scaled, 5-10cm wide leaves spreading to over 1m. Reliable blooming at full size, its distinctive tripinmate inflorescence rises to 120cm. From the thick, dark green scape sprouts, in cylindrical fashion, short, complanate primary and secondary branches to 15cm. The densely imbricate, shiny floral bracts
glow a deep orange-red, contrasting distinctly against the dark greens and the eventual blue petals. Unlike the many other Costa Rican Vrieseas, V. lutheriana more closely resembles the likes of V. elata and V. zamorensis from the Andes. In cultivation this plant requires plenty of space, suiting landscaping, shadehouse and greenhouse environments. With moderate watering and occasional fertilising, its strong leaves can take most extremes, surviving, unblemished, temperatures approaching freezing and exceeding 43deg. at Repton, near Coffs Harbour. The flowering period lasts many months over spring, summer and autumn, ending with a few robust offsets being produced at the base of the plant.

GROWN BY:  Peter Tristram
PHOTOGRAPH BY:  Peter Tristram

VALE: RUTH WILSON

Society members were saddened to hear that Life Member Ruth Wilson has passed away. Ruth was an active member of the Society since joining with husband Bert in the early seventies. Many of our members will remember with fondness the work done by Ruth and Bert in catering for our various shows and other functions. When it seemed that our reserves were running out there was always a warm cup to revive us. Their efforts in catering for our Combined Shows commenced with the first show in 1978 and continued for several years, setting standards that are still being followed. For some years she also staged a mini display in the foyer of our Combined Show. On many occasions Ruth and Bert held garden parties for the Society at their home in Witton Rd., Indooroopilly and our members enjoyed seeing Ruth’s beautiful collection of bromeliads and the wonderful Orchids grown by Bert. In recent years Ruth maintained a reduced collection of plants at their new home in New Farm. Although reduced in numbers the quality was still maintained with Ruth’s plants still winning prizes at our monthly meetings. Ruth and Bert were made life members in recognition of their contribution to the Society.

The Society mourns the loss of a good member and all members extend their condolences to Bert in his bereavement.
Flower Induction and Inhibition in Bromeliads
Peter R. Paroz

One of the unusual attributes of bromeliads is that they can be induced to flower by chemical agents. The earliest references mention the practice of using smoke from fires to induce flowering in glasshouse-grown pineapples in the Azores Islands. In 1932, Rodriquez showed that it was the ethylene component of the smoke which was the active agent. Since that time, there have been many investigations for other active materials. In recent years there have been numerous references in the BSI Journal to the use of A.N.A, Ethepon, and B.O.H. for bromeliad flower induction. In commercial pineapple culture, A.N.A. and Ethrel are used to induce flowering in cultivars of Ananas comosus.

The usefulness of this procedure was limited because flowering could be induced, but there was no way to inhibit the natural flowering of the plant in a manner which allowed flower induction at a later time. In pineapples, A.N.A., which induces flowering at 10 ppm, effectively inhibits flowering at 100 ppm, and the duration of inhibition is unpredictable and uncontrollable.

Research has established that internally produced ethylene is the active agent which triggers the flowering mechanism. Recent studies have identified the biochemical pathways which produce this endogenous ethylene and have opened up the possibility for the complete control of flowering in bromeliads.

A recent paper by DeProft, \textsuperscript{1} and others, demonstrates the potential value of this new information. The test plants for this experiment were one-year-old seedlings of Guzmania lingulata var. minor which were treated by pouring 10ml of solution, at the concentration nominated, into the centre of the plant, and observed, in part, for the time to first flower open and the number of plants which flowered.
**Flower Induction** (cont)

The results of De Proft's experiments are shown in the following table:

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Concentration</th>
<th>% Flowering</th>
<th>First Flower (Days)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td></td>
<td>60</td>
<td>78 +/- 21</td>
</tr>
<tr>
<td>AVG</td>
<td>0.1 mM</td>
<td>0</td>
<td>Infinity</td>
</tr>
<tr>
<td>IAA</td>
<td>0.1 mM</td>
<td>80</td>
<td>75 +/- 20</td>
</tr>
<tr>
<td>ACC</td>
<td>0.4 mM</td>
<td>100</td>
<td>75 +/- 13</td>
</tr>
<tr>
<td>AVG 0.1mM + AVG</td>
<td>0.4 mM</td>
<td>100</td>
<td>76 +/- 9</td>
</tr>
<tr>
<td>Ethephon</td>
<td>500 ppm</td>
<td>100</td>
<td>74 +/- 9</td>
</tr>
</tbody>
</table>

These results show the complete inhibition of flowering by AVG and the induction, or restoration, of flowering by ACC. The other point of note is the substantial reduction in the spread of flowering in the treated plants.

These results have far reaching implications for all who grow bromeliads: the hobbyist, the nurseryman, or pineapple farmer, because now there is the potential to grow plants for flowering to a tight and predetermined schedule.

Interested readers are referred to the original paper for full details of the experiment and results.

**END NOTES:**


2. A.N.A. 1-naphthyl acetic acid
ACC 1-aminoacyclopropene-1-carboxylic acid
AVG aminoethoxyvinyl glycol
B.O.H. 2-hydroxy ethylhydrazine
Ethrel, ethephon 2-chlorethyl phosphonic acid
IAA indole acetic acid

(This article has been reprinted from an earlier issue of "Bromeliaceae")
Tillandsia Watering

Nev Ryan

Tillandsias come from a wide variety of environments such as cloud forest, tropical rainforest, open scrub, deserts and from sea level to over 3000m altitude. We gather a wide variety of these plants into our collections in South East Queensland and try to grow them here. It can be to our benefit, and the plants, to think about the different habitats they originate from, in regards as to how we water them.

My plants are growing under shade cloth, some 50% and some 75% shade. I also grow another group on an east facing wall in full morning sun until about 11.00 am. The plants that are growing in the morning sun are varieties like Tillandsia xerographica, T. ionantha, T. latifolia, T. palacea, T. tectorum, other varieties of silver Tillandsia and some form of T. fasciculata. In winter I water these plants weekly, more often if we get some warm winter days. In summer I water about 3 times a week, unless the weather is very hot or windy, then I would water daily. If you over-water plants like T. palacea, T. tectorum and T. cacticola you encourage algae growth and you lose the silver foliage that is the beauty of the plant.

The plants that I grow under shade cloth are the balance of my collection which contains approximately 300 varieties. The plants in this area that I don’t think need as much water I grow in the top range, that is near the roof. These plants, T. plumosa, T. atroviridipetala, T. crocata, T. cacticola, T. lorentziana, T. magnusiana, T. lepidosepala, are mainly plants from drier conditions. These plants are above the irrigation system and only get watered by hand, which I do weekly all year round. The lower range, that is from about 300mm to about 1.5m below the roof, is watered twice a week in the winter and every second day to daily in the summer, depending on the weather conditions.

As the humidity here can be lower in winter than some Tillandsias need, it is sometimes necessary to water more often. When I water I don’t saturate everything every time. It can be just a walk through the shade house spraying water here and there. This information is for Tillandsias mounted on cork or other medium. As everyone’s growing conditions are different one has to take them into consideration when watering Tillandsias.
Forthcoming Events:

Plant of the Month:
Each month, excepting those months when Mini Shows are conducted, members are invited to table plants from genera starting with consecutive letters of the alphabet for display and commentary.

21st September:
'F' genera, including Fosterella, etc., and 'G' genera, including Guzmania, etc.

19th October:
NO 'Plant of the Month' because of Mini Show

Monthly Meetings:
21st September:
- Popular Vote: Any genus, any species.
- Plant Commentary: P. Hobbs
- Plant of the Month: 'F' and 'G' genera
- 'All about shadecloth' A talk by shadecloth supplier.

19th October:
- Mini Show:
  CLASS 1: Nidularium species & hybrids
  CLASS 2: Tillandsia species & hybrids
  CLASS 3: Hechtia species and hybrids
- Plant commentary on Mini Show by Judges.
- Members Choice show table (non competitive).
- 'Bromeliad Breeding' by Bill Morris.

Study Group:
Meetings: 7:00am, 30th September.
7:00am, 28th October.
PH: (07)33511203

R. H. S. Qld. Inc. Show:
14th & 15th October
A display will be staged by our Society at the Royal Horticultural Society Plant Spectacular Show at the Mt. Coot-tha Botanical Gardens on Saturday 14th and Sunday 15th October.
Members are requested to assist with plants for the display and to act as stewards at the show.

Show Reports:

Popular Vote Results:
17th August
Advanced -
1. Tillandsia carlsoniae L. & O. Trevor
2. Tillandsia gardneri N. Ryan

Intermediate -
1. Tillandsia butzii D. & J. Upton
2. Aechmea orlandiana Ensign

Novice -
1. Tillandsia leonamiana N. Weir
2. Tillandsia Kolbii (tie) P. Crawford
Neoregelia Charm (tie) P. Crawford

Mini Show:
20th July
Advanced -
CLASS 1: Aechmea species and hybrids
1. A. distichantha x chvetnii L. & O. Trevor
2. A. fasciata var. purpurea L. & O. Trevor

CLASS 2: Vriesea (hybrids only)
1. V. Christiane L. & O. Trevor
2. V. Orange Crush L. & O. Trevor
CLASS 3: Pitcairnioideae species and hybrids (not otherwise listed)
   1. Hechtia texensis  D. Andersen
   2. No entry

Intermediate -

CLASS 1:
   1. A. maculata  N. Rowe & G. Alzie wood
   2. A. orlandiana  D. Upton

CLASS 2:
   1. V. saundersii × platynoma f. Albo-Marginata
      N. Rowe & G. Alzie wood
   2. V. Carlsbad
      N. Rowe & G. Alzie wood

CLASS 3:
   1. Deuterocohnia brevifolia  B. Paulsen

Novice -

CLASS 1:
   1. A. orlandiana  N. & L. Weir
   2. A. lueddemanni na  N. & L. Weir

CLASSES 2 & 3. NO ENTRIES

R.N.A. EXHIBITION DISPLAY

The Society staged a display at the RNA Show again this year. The display was in the horticultural pavilion and was attended by stewards for the 10 days that the Show was run. The display included an epiphyte tree as the central theme which was adorned with a good number of plants from several genera including Tillandsia, Vriesea, Neoregelia. Large clumps of T. Usneoides were draped over the branches of the tree providing a very effective balance for the mounted plants. The ground level was filled with a profusion of colourful Neoregelias, Vrieseas, Guzmani as and Aechmeas. The total effect was very colourful and well balanced. The Display Organiser and those members who contributed plants and assistance to put up the display are to be congratulated for the result they achieved. Comments from the public were very complimentary and there were a number of enquiries about the Society and its activities which may lead to new memberships.

New Members:
The Society extends a warm welcome to the following new members:
Ivan and Dawn Hole  Toowoomba.

Trainee Stewards and Judges:
Members interested in becoming Show Stewards and Judges are invited to join the next series of training courses to be conducted in the near future. Further information on these courses may be obtained from:

Olive Trevor  ph: 07-33511203.
Cultural Tips - Hybridising
Odean Head

Hybridising does require some effort and a considerable amount of time before you can fully evaluate the result. It usually takes me 4 or 5 years from the time the seeds are planted to obtain mature plants. You can probably cut at least a year from this schedule by pushing the seedlings with a continuous fertilisation program. You can also do a lot of selection and a considerable amount of culling before the plants mature if you are able to grow them in good light.

All crosses are not successful and to justify our efforts we need to do what we can to increase our odds. This involves selecting the best parents and effective pollination. Be sure to keep good records so that you will know the parentage of any worthwhile hybrids that you make.

PARENT SELECTION: What are we trying to do? Some people have definite goals to develop a plant with a certain color and a good shape. Others are looking for colorful foliage and a beautiful inflorescence or maybe an unusual shape with different colors. The least we can do is to pick two plants with some good qualities and hope that we can pass some good qualities from one plant and combine them with good qualities of the second plant. My efforts thus far have been limited to the plants that are in bloom at the same time. If your objective is to cross plants that bloom at different times, pollen can be saved from the flower of the first plant by freezing it to be used when the second plant blooms.

Most of my experience has been with Neoregelias. The procedures for other genera will vary but the same ‘birds and the bees’ logic will apply.

POLLINATION: In order to make a cross between two plants you must take the pollen from the flower of one plant and apply it to the pistil of the other plant’s flower. Simple enough? Well, it gets a little more involved if you want to make sure that you have actually made a cross between the two plants. First, let’s take a look at the arrangement of the male and female
Cultural Tips - Hybridising (cont)

parts of the flower. The pistil (female part) is located in the center of the flower with the stamens (male parts) surrounding it. The pollen is released by the anthers which are located at the tips of the stamens. To fertilise the flower the pollen must be placed and adhere to the stigma which is located at the tip of the pistil. The stigma is only receptive to pollen for a short period of time while the flower is fully open (usually about mid morning). You may have to apply pollen more than once during this time to increase the chances for conception. In most Neoregelias the stigma is on the same level or above the anthers which keeps them from self pollinating. However, if you are applying pollen from another flower there is a good chance that you will stir up the pollen on the targeted flower causing it to self pollinate. To avert this you can emasculate the targeted flower by carefully removing its stamens before the pollen ripens (usually early morning or the night before the flower fully opens). The pollination can be made with something small, a brush or even a toothpick, that the pollen will adhere to and can be easily transferred to the small pistil. You may want to make multiple crosses to the same plant while it is blooming. Make only one cross per bloom and identify its pollen parent by placing a small label in the pollinated flower.

If the pollination is successful, seeds will begin to develop in the ovary beneath the flower and mature in about 3 months.

(The author's article 'Cultural Tips - Growing Bromeliads from Seed' was printed in the July-August, 1995 issue of "Bromeliaceae" and provides the author's methods for maturing, gathering, planting the seeds, and the growing of the seedlings. Ed)
Collecting in Costa Rica - Vrieseas (Werauhias), Tillandsias and Guzmanias galore!
Peter Tristram

The Central Valley of Costa Rica, flanked by ageless active volcanoes and the mountains of a now almost swallowed tectonic plate, is largely cleared and densely settled. Large areas of forest still exist however, in the south along the slopes of the Cordillera de Talamanca. Unlike much of Costa Rica, this mountain range is not of volcanic origin, extending way south into Panama.

One area in which Chester Skotak and I collected was at the northern end of the Talamancas, overlooked to the north by the huge, rounded cone of Volcano Irazu which last erupted in the 1960’s, coating the countryside in fertile black ash and spewing lava flows almost to the city of Cartago, blocking the Pan American highway. The area bore few scars of this event, however the huge mound of Irazu and the shattered remains of a never completed cathedral bore testament to the fractured nature of the Earth’s crust in that part of the world.

The Rio Grande de Orosi has its source high in the Talamancas and few, if any, roads extend past the hydro-electric stations and the Tapanti National Park at the top end of the valley. Along the lower valley little original vegetation remained, and most was on private property! Trees, gardens and hedges full of large Tillandsia fasciculata, T. variabilis, T. oerstediana, T. utriculata and smaller T. bulbosa, T. tricolor, T. juncea, T. leiboldiana the like abounded as did Guzmania lingulata and G. monostachia in various areas.

Not much collecting was attempted until we arrived at the narrow end of the valley where the road traversed the Orosi. In trees along the road a cocktail of Tillandsias, Racinaea, Vrieseas (including Werauhias) and Guzmanias were to be found. A beautiful discolor form of Tillandsia complanata were lined up in rows along the sunny side of some branches, while Racinaea spiculosa and R. adpressa choked others. In a tree fork a clump of boldly striped Guzmania was seen, so the old Toyota truck was backed up and a
Collecting in Costa Rica (cont)

few specimens finally dislodged with the collecting pole. This has since been named Guzmania skotakii, a new species. The area was profuse with other Guzmania species but most were ‘cone-heads’ - a typically flowering group of horticulturally uninteresting species.

We drove back down the valley a few kilometres to an hydroelectric station access road and started a steep ascent. In the many she-oaks (Casuarina species) along property boundaries were more Tillandsia complanata as well as red-blotched rosettes of T. excelsa of all sizes, large bulbs of T. butzii, T. punctulata, T. juncea and inflated bulbs of the pretty Vriesea castano-bulbosa with its leathery, silver-frosted leaves, ping inflorescence and green flowers making a distinctive contrast on the drier side of trees and fence posts. These were the tree-top survivors of the rainforest which blanketed this area until fairly recently. The farms were primarily coffee being at 1000-1200m altitude, with dairy cattle a little higher up.

Continuing climbing, the vegetation thickened as the cloud base approached. It was there, at 1500m, that Guzmanias and Vrieseas (especially Werauhia) reached epiphytic dominance. All levels of vegetation were festooned with an enormous variety of Tillandsioideae based species, most of which were not seen at lower altitudes. In this life zone, Tillandsia excelsa (large form), Vriesea castano-bulbosa, Racinaea spiculosa and R. adpressa were still prevalent, but the others had disappeared. It was the domain of the ‘Thecophyllloid’ Vrieseas, now lumped into the new genus Werauhia, especially in sunny and more exposed sites and the Guzmanias dominated the damper and shadier places. Vriesea leucophylla with its decurved, dazzling pink spike and maroon crossbanded silver-frosted triangular leaves were everywhere in almost every niche, from full sun to total shade, from ground level, by the shiny, yellow and green, crossbanded rosettes of V. nephrolepis, many displaying their typical large, secund, polystichous, yellow flowers and hooting maroon primary bracts. (Incidentally, further up these gave way to V. ouroensis with broader, flatter rosettes and bright cross-banding as the UV level increased and a whole new range of predominantly Vriesea species (We did not collect there).

It was within the protection of the forest that the Guzmanias reached
Collecting in Costa Rica (cont)

dominance, abounding in all shapes, sized and colours. Spectacular red and yellow heads of a large form of Guzmania blassii shone in the dim light and luminous, yellow flowered, red hot pokers of a small pin-striped Guzmania vaguely similar to G. nicaraguensis hung along the undersides of low branches, their bases submerged in moss. Long clumping strands of G. angustifolia hung and climbed among the varied stems along the rainforest margin, occasional clumps sporting bright red ‘noses’ as they weaved among the orchids, anthuriums, philodendrons and epiphytic ferns. The new G. skotakii again found as was a selection of ‘cone-heads’.

In this area, Chester Skotak assured me that the beautiful Aechmea veitchii could be found. He pointed me towards the deepest shade following a precipitous gorge, where few bromeliads, excepting stems of climbing Pitcairnias, could survive. About thirty metres down, spectacularly in bloom, the straggly, long-leaved rosettes were eventually seen, sprawling between huge rocks, rhizomes hidden in the decaying vegetation. Finally a few specimens were obtained, the rest left to propagate.

Back in the sun, the moss along the forest margin contained a profusion of seedlings of many genera, making good grazing for the local cows. Large, broad-leaved, purple-black rosettes of Vriesea latissima made a distinctive colour contrast in terrestrial areas. The older trees here were festooned with epiphytes to the extent that the ground under them was littered with fallen tree-branch gardens in various stages of decay. In these ‘gardens’, species such as Tillandsia longifolia Racinaea spiculosa were found as well as tight bulbs of Vriesea castano-bulbosa, Racinaea contorta, R. adpressa, Vriesea tonduziana and a host of unidentifiable seedlings. One significant find was Tillandsia cauliflora, an unusual small Tillandsia with bright vermilion, white-dusted tubes and a stunning cylindrical inflorescence. It proved extremely difficult to collect.
Collecting in Costa Rica (cont)

sheltering fork, often flanked by clumps of Guzmania condensata with their long, purple-pointed, yellow-green leaves and orange and yellow branched inflorescences. M. pleiosticha was a true giant standing around 1 metre high and spreading 2 metres, making collection impossible! Fortunately, in the terrestrial moss gardens, occasional seedlings could be found, identifiable by their leaf texture and dark sheaths.

Beside a stand of spectacular, prickly, broadly-lobed treeferns adjoining a nearby Cyprus 'reafforestation' area, the unmistakable leaves of Guzmania plicatifolia were noticed in the shade, looking all the world like clumps of palm seedlings with their multi-channelled, long, thin, arching leaves. In the open, atop a large tree stump, a dazzling, zigzag, pink spray belonged to a huge Tillandsia longifolia, flowering for the first and last time, a natural occurrence unlike the fate of the tree from which it certainly fell.

Everywhere, in every niche, were a myriad of plant species, especially bromeliads, many of which were unknown to me. Most of the plants collected in this area easily survived the rigours of transport and quarantine, flourishing in my nursery today, many still remaining unidentified.

Neoregelia monstrosum? N. hatschbachii? N. cruenta (red form)!
John Higgins

I originally obtained this plant as Neoregelia monstrosum from a local grower in Queensland. When Harry Luther attended Bromeliads III in Brisbane in 1983 he tentatively identified this plant as N. hatschbachii in the Brisbane Botanical Gardens. At the time Harry did not have the benefit of close inspection and his identification was given on the run during a whirlwind tour of the gardens. At Bromeliads 8 earlier this year Derek Butcher advised the conference that further consideration of this plant has led Harry to agree that it is really a red form of N. cruenta.
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